

International Conference of the Genetics Society of Korea

Oct. 20, 2023

MH-UMIseq: improvement of signal-to-noise ratio in NGS of microhaplotype with **unique molecular identifiers**

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Main challenges in forensic DNA profiling

- Main topics of our research group

1



Degraded DNA

- Short amplicon
- 25-plex A-STR / 24-plex Y-STR NGS panel



2



Low copy number DNA (Ongoing study)

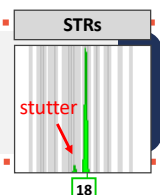
- PTA-based whole genome amplification (WGA)
- Hybrid-capture for NGS

3



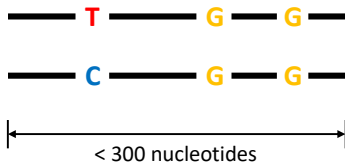
Mixture deconvolution (Ongoing study)

- Microhaplotypes NGS



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Microhaplotypes



Next generation sequencing (NGS)

- High throughput multiplexing
- Phase-known haplotyping

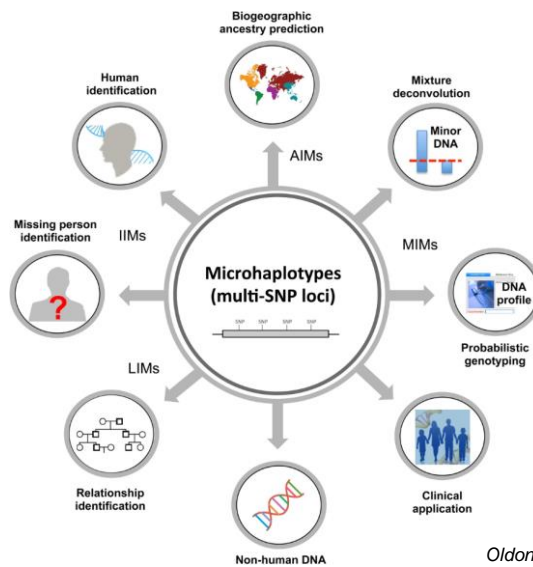


Genotype : TGG / CGG

- Low mutation rate
- High heterozygosity
- No stutter artifacts

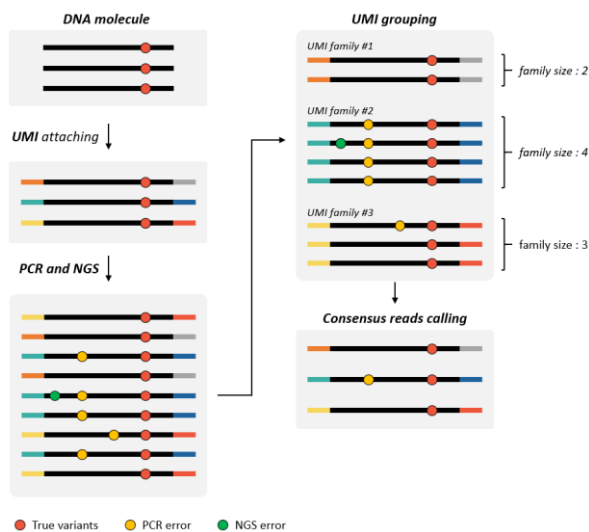
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Microhaplotypes



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Unique molecular identifiers (UMIs)



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Attaching UMIs to DNA molecules

1. PCR-based method



1. Multiplex PCR

- **Pros** : High sensitivity (≈ 1 ng)
- **Cons** : Small number of targets
(Difficult to design primer)



Suitable for forensic DNA

2. Adapter ligation method



- **Pros** : Large-scale multiplexing
(> 100 kb)
- **Cons** : Low sensitivity (> 10 ng)

e.g. QIAseq Targeted DNA Panel (Qiagen)

Allele- / locus- drop out, < 2 ng DNA

Truelsen (2021) Sci. Rep.

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Research objectives

General objective

Construct **UMI-based amplicon sequencing system for microhaplotype** and investigate **false-positive rate** of UMI data according to **the amount of input DNA**.

Specific objectives

1. Development of **MH-UMIseq Panel**
2. Construction of **MH-UMI data analysis pipeline**
3. **Evaluating performance** of MH-UMIseq

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1. Development of MH-UMIseq Panel

Marker selection

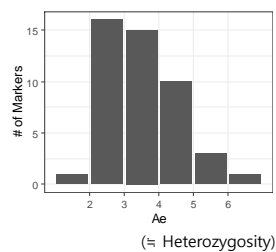
130 Microhaplotypes



Kidd (2017) FSIG

46 Microhaplotypes

- Consist of 155 SNPs
- Target size: 124 ~ 249 bp

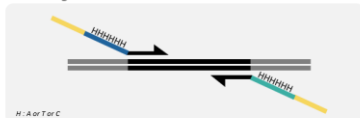


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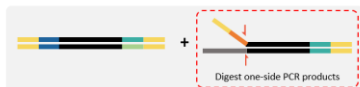
1. Development of MH-UMIseq Panel

MH-UMIseq

1. Barcoding PCR (x 3 cycles)



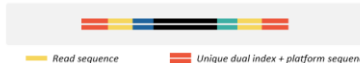
2. Nuclease reaction



3. Boosting PCR



4. Indexing PCR



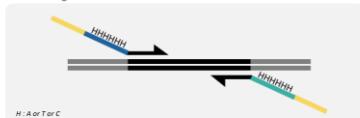
- **UMI barcode**
 - $\text{Poly}(H_6) \times \text{Poly}(H_6) = \text{Poly}(H_{12})$
 - = $3^{12} = 531 \text{ K}$
- **Remove PCR artifacts by Nuclease**
by *CleanPlex (Paragon Genomics)*
- **UMI-attached target amplification by Boosting PCR**

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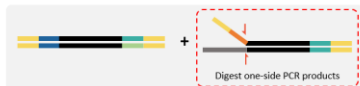
1. Development of MH-UMIseq Panel

MH-UMIseq

1. Barcoding PCR (x 3 cycles)



2. Nuclease reaction



3. Boosting PCR



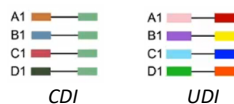
4. Indexing PCR



➤ To mitigate PCR and NGS error

- **High-fidelity enzyme (> x 300)**
- **Unique dual index (UDI)**

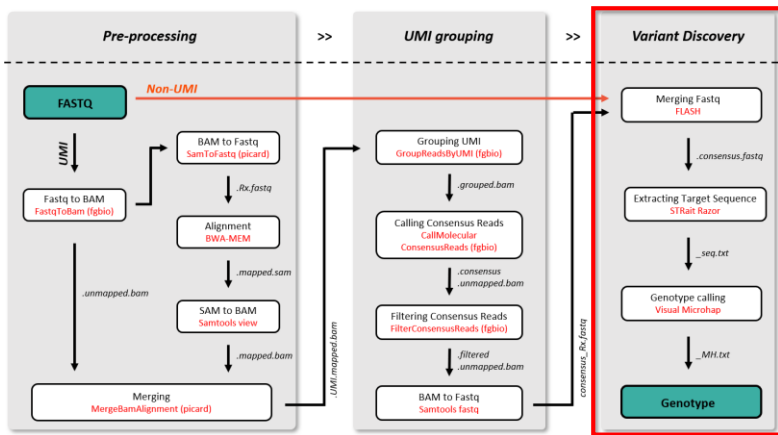
↔ Combinatorial dual index (CDI)



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2. Construction of MH-UMI data analysis pipeline

- Linux-based pipeline + *STRait Razor* + *Visual Microhap*



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2. Construction of MH-UMI data analysis pipeline

Visual Microhap (v1.1) for MPS Data by STRait Razor 3.0

Marker Information

File: MH56_Ref_Info.txt

56 markers: Loading completed!

Analysis Option

- Minimum read count: 100
- Background noise level: 0.05
- Homopolymer (> 7) error (Indel) followed by '@' in Cryptic Var: 0.15
- Allele Proportion (AP): 0.25
 - 'c': in column AP
 - Exclude in result

[Download demo file](#) (Microhaplotyping panel v2.0)

[Marker Information for 56 Microhaps](#)

[Microhap Sequence Data by STRait Razor 3.0](#)

[Download](#) [Quick Start Guide](#)

Sequence-based Data

File: MH56-2800M_Seq.txt

56 markers: Analysis completed!

Marker.No	Chr-Pos	Reference	Obs.	Cryptic Var.	Read Valid	Gross AP
mh01KK_001:1	chr1:3826568	rs4648344T>rs6683840G>rs58111155G	CAG	3826587G>A	3128 51.5 (28.3)	
mh01KK_001:2	chr1:3826568	rs4648344T>rs6683840G>rs58111155G	CAG		2950 48.5 (26.6)	
mh01KK_002:1	chr1:216461086	rs4528199G rs6604596A	GA		2967 55.9 (47.6)	
mh01KK_002:2	chr1:216461086	rs4528199G>rs6604596A	AA		2338 44.1 (37.5)	
mh01KK_106:1	chr1:4167404	rs12123330C rs16840876A rs56212601G rs4468133G>	CAGA		5208 100.0 (70.0)	
mh01KK_117:1	chr1:204664212	rs17413714A>	CACC		6671 100.0 (61.7)	

[Download Results](#)

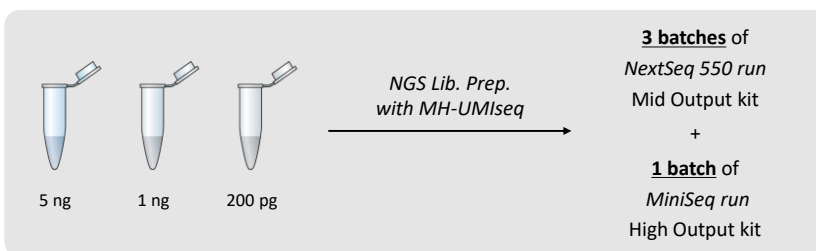
Kwon (2022) FSIG

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3. Evaluating performance of MH-UMIseq

Material and Method

- 31 Korean blood samples (20 Male + 11 Female) → *Real-time PCR quantification*
Severance IRB. 4-2022-1534
- 31 samples x 3 types of input DNA = 93 samples

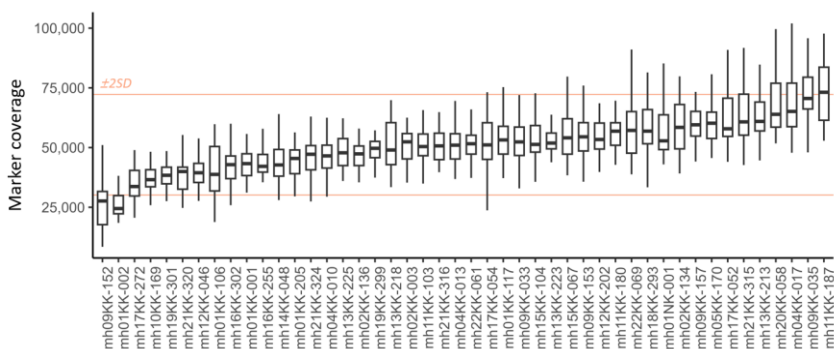


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3. Evaluating performance of MH-UMIseq

Marker coverage

e.g. 1ng of DNA

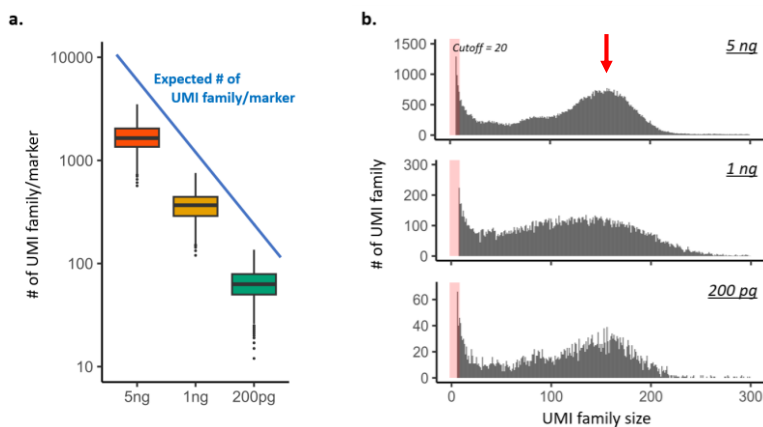


(Manuscript in preparation)

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3. Evaluating performance of MH-UMIseq

UMI family distribution



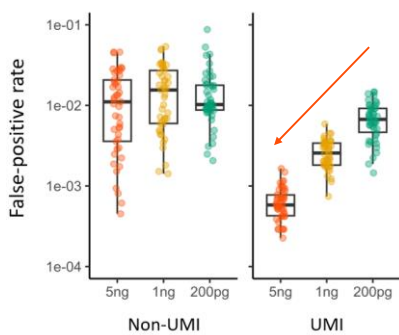
\therefore # of UMI family \propto amount of input DNA

(Manuscript in preparation)

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3. Evaluating performance of MH-UMIseq

False-positive rate



➤ Based on Minimum analytical options

UMI type	Min. read cov. / UMI family size	noise cutoff
Non-UMI	x 20	0.001
UMI	x 20	0.000

\therefore **False-positive rate: non-UMI > UMI**

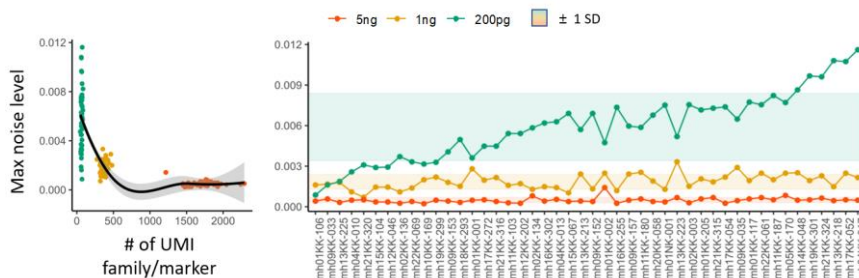
\therefore **amount of input DNA**
 \propto efficiency of UMI system

(Manuscript in preparation)

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3. Evaluating performance of MH-UMIseq

Max noise level of MH-UMIseq



➤ Expected resolution of MH-UMIseq

	5ng	1ng	200pg
Minor component ratio	1:231	1:64	1:13

(Manuscript in preparation)

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Summary

- ◆ The UMI-based 46-plex amplicon sequencing system and UMI data analysis pipeline for microhaplotypes were well established.
- ◆ The false-positive rate was significantly reduced in UMI system, especially for higher amounts of input DNA.
- ◆ The noise of MH-UMIseq is mainly affected by the number of molecules (= amount of input DNA).
 - The noise level decreases as the input increases.
- ◆ To obtain a resolution of 1:50 or higher, ≥ 1 ng of DNA must be used.

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Future application of UMI system

- ◆ It is expected to achieve high accuracy and sensitivity (resolution) in **mixed DNA analysis**, especially for high amount of DNA.

+ DNA Methylation analysis

- ◆ It is expected to improve the accuracy and sensitivity of **age, body fluid prediction** by **DNA methylation analysis**.
- ◆ It enables simultaneous **human identification** and **methylation analysis** of **mixed samples**.

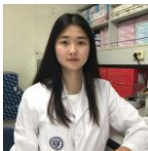
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Acknowledgements

➤ Yonsei DNA profiling group



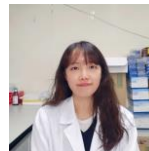
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This work was supported by the Korean National Police Agency
(Project Number: 0411-20230029)

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