

Massively Parallel Sequencing Analysis of Whole Mitochondrial Genomes in Hair Shaft, Blood and Buccal Swab Samples

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1. Introduction



CHALLENGING

- Hair shafts
- Skeletal remains

REFERENCE

- Buccal swab
- Blood



1. Introduction



CHALLENGING

- Hair shafts
- Skeletal remains

- Insufficient amount of available DNA
- High level of DNA fragmentation
- ➔ **Small amplicon strategy**

J. Forensic Sci. 2001 Mar;46(2):247-53.
Improved mtDNA sequence analysis of forensic remains using a "mini-primer set" amplification strategy.
 Gabriel MM¹, Huffine EE, Ryan JH, Holland MM, Parsons TJ.
 Int J Legal Med (2008) 122:385-388
 DOI 10.1007/s00414-008-0227-5

Author info
 1. mtDNA Seq

ORIGINAL ARTICLE

'Mitominis': multiplex PCR analysis of reduced size amplicons for compound sequence analysis of the entire mtDNA control region in highly degraded samples

Cordula Eichmann · Walther Parson



1. Introduction

- **MPS strategy for challenging sample type**

Precision ID mtDNA Panels

Mitochondrial DNA Genome
16,569bp

16024 16365 73 340 438 576

HV1 HV2 HV3

16569/1

1 2 3 4 5 6 7 8 9 10 11 12 13 14

16569/1

ThermoFisher SCIENTIFIC

Precision ID mtDNA whole Genome Panel

- 163 bp average amplicon length
- 81 primer pairs
- 2-pool multiplex assay
- Degraded and compromised sample

Precision ID mtDNA Control Region Panel

- 153 bp average amplicon length
- 7 primer pairs

1. Introduction

REFERENCE

- Buccal swab
- Blood



- Often taken directly from a suspect
- Low level of DNA fragmentation
- Impacts of NUMTs (*nuclear mitochond*
- ➔ **Long-range amplification strategy**

Forensic Science International: Genetics 13 (2014) 128–135
Contents lists available at ScienceDirect
Forensic Science International: Genetics
journal homepage: www.elsevier.com/locate/fsig

Forensic Science International: Genetics 12 (2014) 128–135
Contents lists available at ScienceDirect
Forensic Science International: Genetics
journal homepage: www.elsevier.com/locate/fsig

High-quality and high-throughput massively parallel sequencing of the human mitochondrial genome using the Illumina MiSeq
Jonathan L. King^{a,1,*}, Bobby L. LaRue^{a,1}, Nicole M. Novroski^a, Monika Stolarova^a, Seung Bum Seo^a, Xiangpei Zeng^a, David H. Warshawer^a, Carey P. Davis^a, Walther Parson^{b,c}, Anhi Sajatnia^a, Bruce Budowle^{a,d}

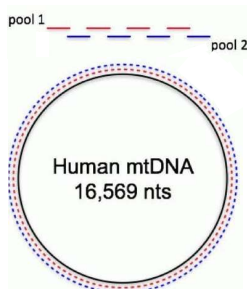
^aInstitute of Applied Genetics, Department of Molecular and Medical Genetics, University of North Texas Health Science Center, 3500 Camp Bowie Blvd., Fort Worth, TX 76107, USA
^bInstitute of Legal Medicine, Innsbruck Medical University, Innsbruck, Austria
^cParis Lodron University of Salzburg, University Park, A-5020
^dDepartment of Forensic Medicine, Rush Medical College, 1725 East Chicago Avenue, Chicago, Illinois, United States
^eCenter of Excellence in Genetic Medicine Research (CEGM), King Abdulaziz University, Jeddah, Saudi Arabia



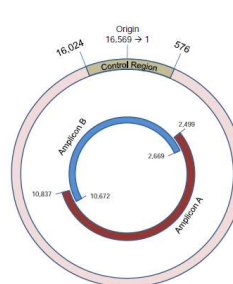
1. Introduction

➤ MPS strategy for forensic sample type

[Hair shafts]



[Reference]



Precision ID mtDNA Whole Genome Panel

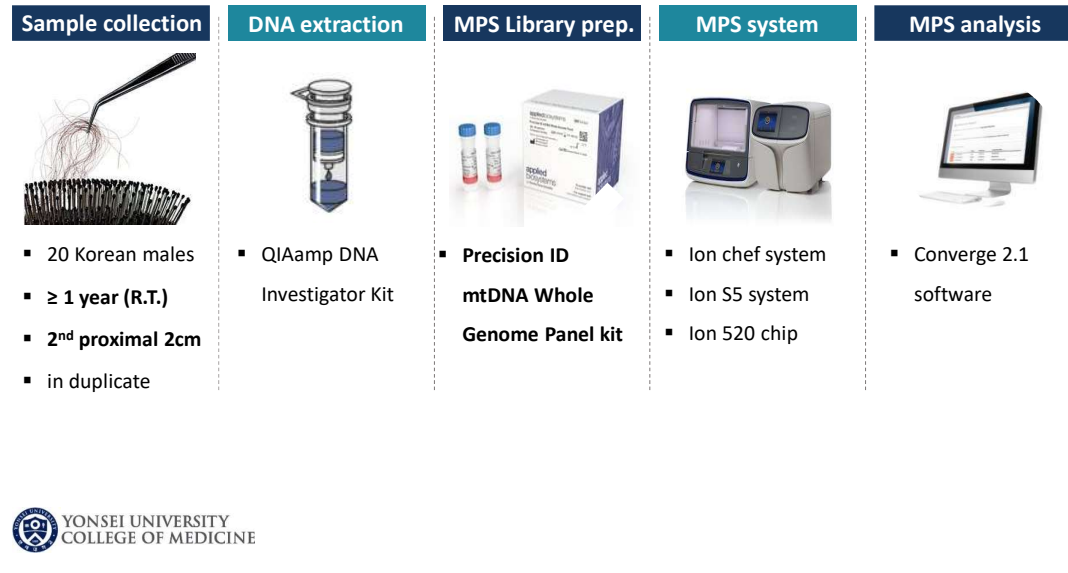
- 163 bp average amplicon length
- 81 primer pairs
- 2-pool multiplex assay
- Degraded and compromised sample

Long-range amplification

- 8,500 bp average amplicon length
- 2 primer pairs
- Avoiding impacts of NUMTs
- Intact and high-quality sample

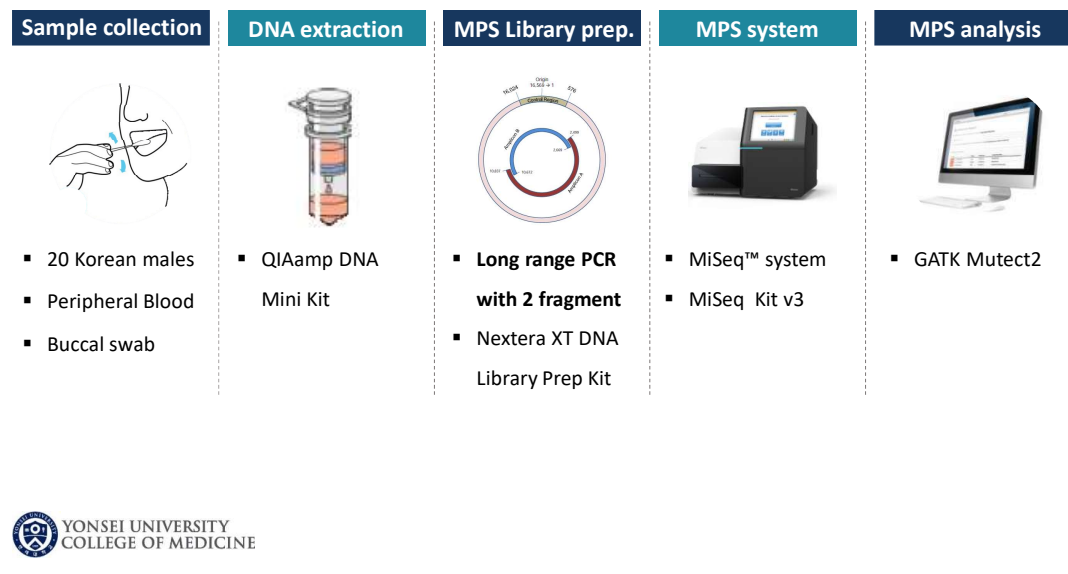
2. Materials & Methods

➤ MPS workflow of hair shafts



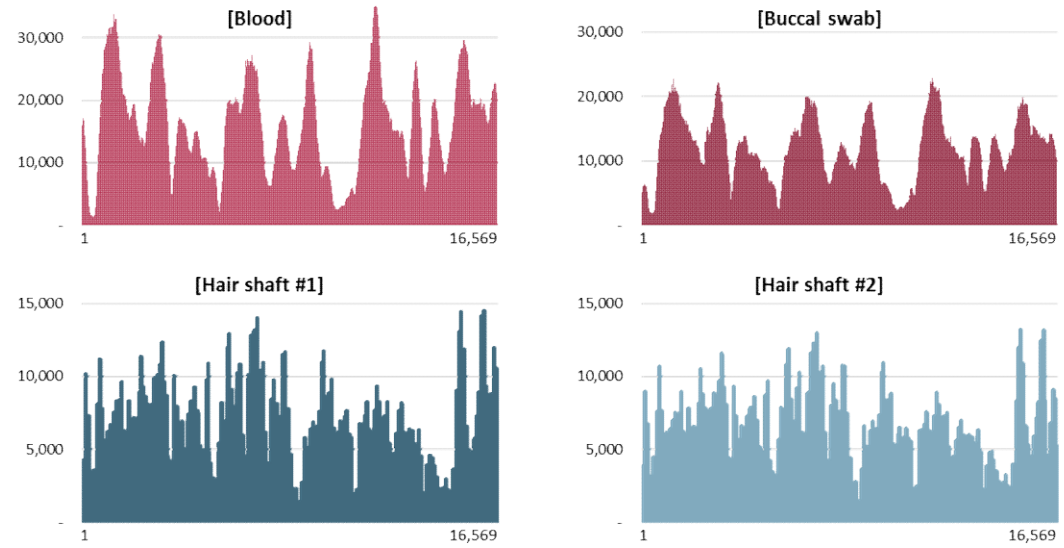
2. Materials & Methods

➤ MPS workflow of reference samples



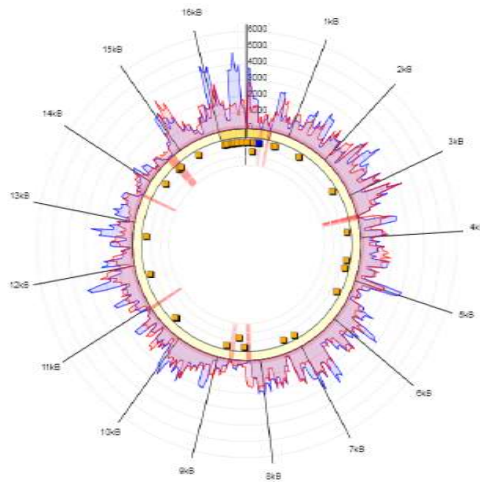
3. Results

➤ MPS coverage



3. Results

➤ MPS data analysis of hair shafts using **Converge software**



[Circular coverage plot]

- Forward coverage
- Reverse coverage
- Deletion
- Insertion
- Mutation




3. Results

➤ MPS data analysis of hair shafts using **Converge software**

[Converge software result]

Sample	3001_Hlon/press_085		Close haplogroup Y1																								
Regions	-16569																										
Position	Ref	SampleVariant	Var Freq	Type	read Coverage	read Coverage	Allele Coverage	Allele Coverage	Coverage	Coverage	CV%	AN	T%	C%	N%	ins%	del%	Polymorphism	Control	Region	State	Frequency	Artefact	Far Strand	Bleed	Strand Bi	EMPOP
73	A	G	G	99.8	SNP	3616	2865	6466	3607	2859	99.8	0.2	0	0	0	0.1	0	0	73G	73G	confirmed	99.8	True variant	0.5	0.6	unchecked	
146	T	C	C	98.6	SNP	1885	1378	3218	1858	1360	0	0	0.1	98.6	0	0.6	1.3	146C	146C	confirmed	98.6	True variant	0.5	0.6	unchecked		
152	T	T	Y	34.6	SNP	1883	1393	1132	649	483	0	0	65.4	34.6	0	1.2	0	152Y	152Y	possible	34.6	Point Heteroplasmy	0.5	0.6	unchecked		
263	A	G	G	99.9	SNP	433	438	870	432	438	99.9	0	0	0	0	0.6	0.1	263G	263G	confirmed	99.9	True variant	0.5	0.5	unchecked		
309	T	+	C	42.5	INS	365	798	494	171	323	0	0	0	0	0	54.9	0	309.1C	309.1C	likely	42.5	Length Heteroplasmy	0.5	0.7	confirmed		
315	G	+	C	75.6	INS	357	802	876	225	651	0	0	0	0	0	85.7	0	315.1C	315.1C	likely	75.6	True variant	0.6	0.7	confirmed		
709	G	A	A	98.6	SNP	3087	2871	5874	3040	2834	1.3	98.6	0	0	0	1.4	0.1	709A		confirmed	98.6	True variant	0.5	0.5	unchecked		
750	A	G	G	99.4	SNP	791	1842	2618	789	1829	99.4	0	0	0	0	0.7	0.5	750G		confirmed	99.4	True variant	0.5	0.7	unchecked		
1438	A	G	G	99.5	SNP	1186	709	1885	1185	700	99.5	0.5	0	0	0	0.4	0	1438G		confirmed	99.5	True variant	0.5	0.6	unchecked		
2706	A	G	G	98.6	SNP	1644	1486	3086	1617	1469	98.6	1.4	0	0	0	0.6	0	2706G		confirmed	98.6	True variant	0.5	0.5	unchecked		
3834	G	A	A	100	SNP	303	696	999	303	696	0	100	0	0	0	0	0	3834A		confirmed	100	True variant	0.5	0.7	unchecked		
4562	A	G	G	97.3	SNP	1771	1386	3071	1710	1361	97.3	2.2	0	0	0	0.3	0.5	4562G		confirmed	97.3	True variant	0.5	0.6	unexpected		
4769	A	G	G	99	SNP	1013	1136	2127	1011	1116	99	0	0	0	0	0.5	1	4769G		confirmed	99	True variant	0.5	0.5	unchecked		
5417	G	A	A	99.8	SNP	176	441	616	175	441	0.2	99.8	0	0	0	1.3	0	5417A		confirmed	99.8	True variant	0.5	0.7	unchecked		
7028	C	T	T	97.9	SNP	2217	1001	3149	2170	979	0	0	97.9	2.1	0	0.1	0	7028T		confirmed	97.9	True variant	0.5	0.7	unchecked		
7325	A	G	G	99.6	SNP	1764	1952	3701	1762	1939	99.6	0.1	0	0	0	0.2	0.3	7325G		confirmed	99.6	True variant	0.5	0.5	unexpected		
8392	G	A	A	99.1	SNP	167	388	550	164	388	0.9	99.1	0	0	0	0.2	0	8392A		confirmed	99.1	True variant	0.5	0.7	unchecked		
8555	T	T	Y	25.1	SNP	791	552	337	208	129	0	0	74.9	25.1	0	0.3	0	8555Y		unclear	25.1	Point Heteroplasmy	0.5	0.6	unexpected		
8860	A	G	G	99.9	SNP	429	840	1276	428	848	99.9	0.1	0	0	0	0.2	0	8860G		confirmed	99.9	True variant	0.5	0.7	unchecked		
10379	A	G	G	97	SNP	953	1527	2406	909	1497	97	2.9	0	0	0	0.3	0.1	10379G		confirmed	97	True variant	0.5	0.6	unchecked		
10398	A	G	G	99	SNP	1073	1746	2790	1061	1729	99	1	0	0	0	0	0	10398G		confirmed	99	True variant	0.5	0.6	unchecked		
11719	G	A	A	99.1	SNP	3040	1028	4032	3019	1013	0.9	99.1	0	0	0	0.3	0	11719A		confirmed	99.1	True variant	0.5	0.7	unchecked		
12705	C	T	T	98.1	SNP	1985	564	2500	1947	553	0	0	98.1	1.9	0	0.2	0	12705T		confirmed	98.1	True variant	0.5	0.8	unchecked		




3. Results

➤ MPS data analysis of hair shafts using **Converge software**

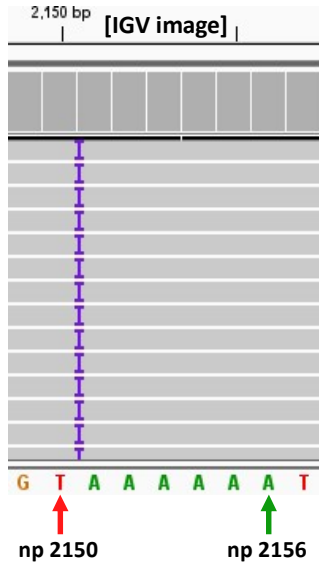
[Converge software result]

Sample	3001_Hlon/press_085		Close haplogroup Y1																								
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263	A	G	G	99.9	SNP	433	438	870	432	438	99.9	0	0	0	0	0.6	0.1	263G	263G	confirmed	99.9	True variant	0.5	0.5	unchecked		
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11719	G	A	A	99.1	SNP	3040	1028	4032	3019	1013	0.9	99.1	0	0	0	0.3	0	11719A		confirmed	99.1	True variant	0.5	0.7	unchecked		
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3. Results

➤ MPS data analysis of hair shafts using **Converge software**



[Converge 2.1 result]

Position	Ref	Sample	Variant	Type	...	Polymorphism
2156	T	+	A	INS	...	2156.1A

→ Automatically calling according to forensic nomenclature (ISFG guidelines)

[GATK Mutect2 result]

#Chrom	Position	Ref	Alt
chrM	2150	T	TA

3. Results

➤ Observed point heteroplasmy of whole mtGenomes

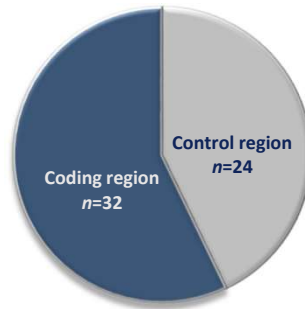
Sample ID	rCRS Position	rCRS Nucleotide	Hair shaft#1	Hair shaft#2	Blood	Buccal swab
Sample 001	152	T	C	T/C(34.6)	T/C(22.0)	C/T(33.4)
	3834	G	A/G(5.4)	A	A	A
	8555	T	T	T/T(25.1)	T	T
	15364	C	C/T(10.4)	C	C	C
	16266	C	T/C(5.6)	T	T	T
Sample 004	16319	G	A/G(8.3)	A/G(7.8)	A	A
	709	G	A/G(41.8)	G	G	G
	16320	C	C/T(10.1)	C	C/T(24.9)	C/T(16.8)
Sample 006	41	C	C/T(20.7)	C	C	C
	11847	G	G/A(13.6)	G	G	G
Sample 007	16150	C	T	T	T	T/C(5.8)
Sample 008	15373	A	A	A/G(32.2)	A	A
	16362	T	C	C/T(17.7)	C	C
Sample 009	8517	G	G	G	G/G(6.4)	G
	16187	C	T	T/C(7.5)	T	T
	11969	G	A/G(5.7)	A	A	A
Sample 012	16192	C	T/C(7.0)	T	T	T
	16311	T	C	C/T(45.2)	C	C
Sample 014	8473	T	C/T(19.2)	C/T(14.3)	C	C
	16362	T	C/T(6.4)	C/T(5.8)	C	C
Sample 015	204	T	T	T	T/C(13.6)	T/C(49.0)
	499	G	A/G(7.6)	A	A	A
	2831	G	A/G(5.7)	A	A	A
	4820	G	A/G(12.9)	A/G(8.0)	A	A
	15034	A	G/A(5.9)	A/G(29.9)	A/G(15.6)	A/G(9.9)
	15236	A	G/A(5.7)	G	G	G
Sample 016	14016	G	A/G(11.4)	A/G(10.3)	A	A
	930	G	A/G(23.0)	A/G(30.9)	A/G(28.3)	A/G(23.0)
Sample 017	15279	T	T	T/C(33.1)	T	T
	16103	A	A	A/G(33.0)	G/A(19.1)	G/A(29.7)
Sample 018	15262	T	C/T(27.1)	C/T(18.4)	C	C
Sample 019	9947	G	G/A(45.6)	G	G	G
	10644	G	G/A(37.3)	G	G/A(18.6)	G/A(19.3)
Sample 020	1200	G	G	G/A(40.0)	G	G
	16162	A	A/G(13.6)	A	A	A

3. Results

➤ Observed point heteroplasmy of whole mtGenomes

- ✓ Variant calling threshold >100X
- ✓ Point heteroplasmy threshold > 400X
- ✓ Minor nucleotides of PHP were observed in > 5% of the total coverage

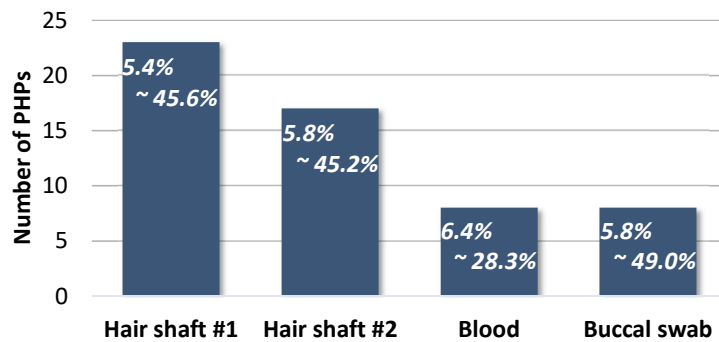
- Total 56 PHPs
- 15/20 donors across 4 tissue sets
- PHPs per donor up to 5,3,2,2 in hair shaft #1, hair shaft#2, blood, buccal swab
- Control region : Coding region = 24 : 32



3. Results

➤ Observed point heteroplasmy of whole mtGenomes

Sample type	Total	Hair shaft #1	Hair shaft #2	Blood	Buccal swab
No. of PHPs	56	23	17	8	8
Proportion range(%)	5.4~49.0	5.4~45.6	5.8~45.2	6.4~28.3	5.8~49.0



4. Discussions

➤ Inverted major nucleotide according to the tissue type

Sample ID	rCRS position	rCRS nucleotide	Hair shaft #1	Hair shaft #2	Blood	Buccal swab
Sample 001	152	T	C	T/c(34.6)	T/c(22.0)	C/t(33.4)
Sample 004	709	G	A/G(41.8)	G	G	G
Sample 015	15034	A	G/A(5.9)	A/G(29.9)	A/G(15.6)	A/G(9.9)
Sample 017	16103	A	A	A/G(33.0)	G/A(19.1)	G/A(29.7)

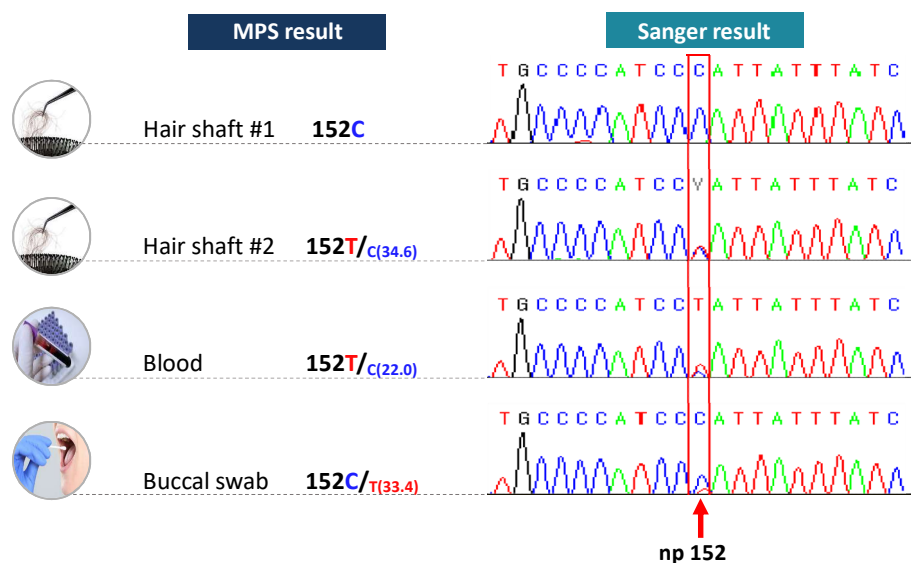
*Minor variants are indicated using smaller font size with their proportion stated in parentheses.



4. Discussions

➤ Inverted major nucleotide according to the tissue type

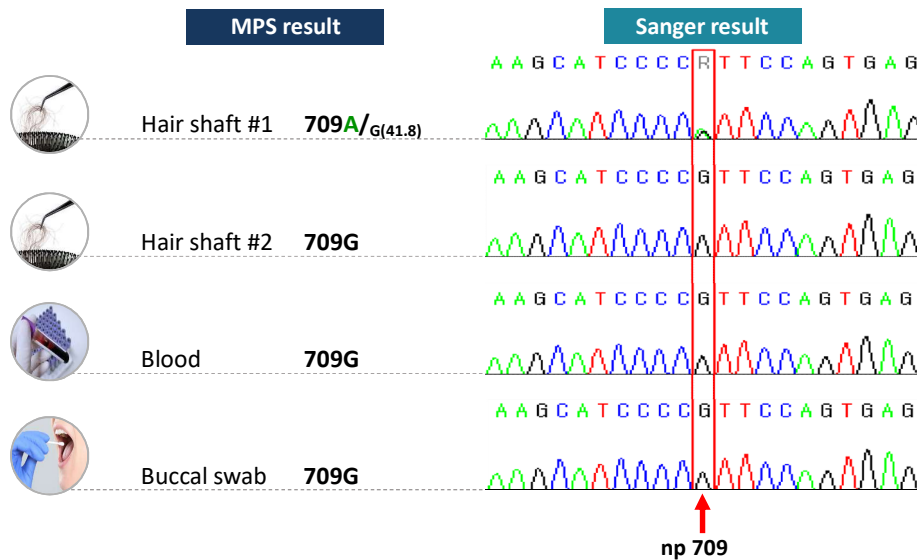
[Sanger sequencing results of *Sample 001*]



4. Discussions

➤ Inverted major nucleotide according to the tissue type

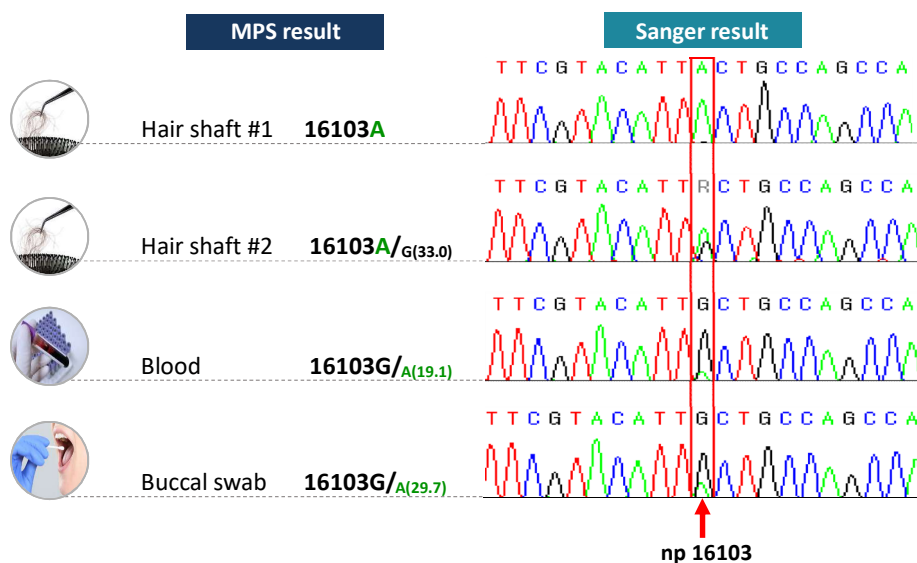
[Sanger sequencing results of *Sample 004*]



4. Discussions

➤ Inverted major nucleotide according to the tissue type

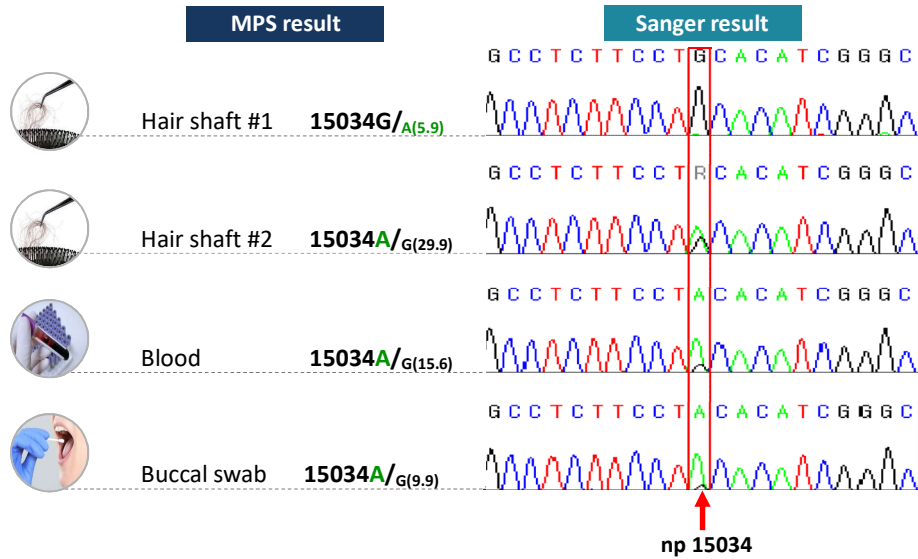
[Sanger sequencing results of *Sample 017*]



4. Discussions

➤ Inverted major nucleotide according to the tissue type

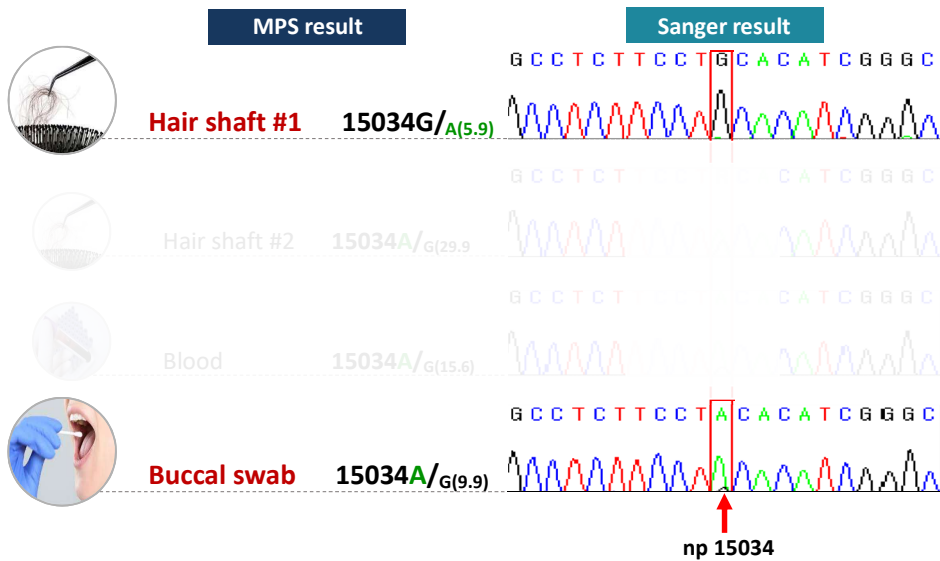
[Sanger sequencing results of *Sample 015*]



4. Discussions

➤ Inverted major nucleotide according to the tissue type

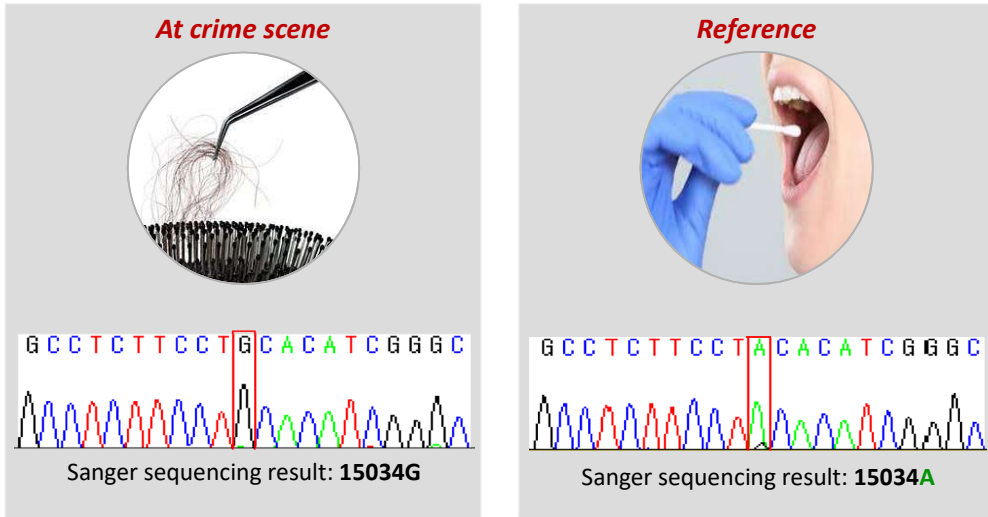
[Sanger sequencing results of *Sample 015*]



4. Discussions

What if..?

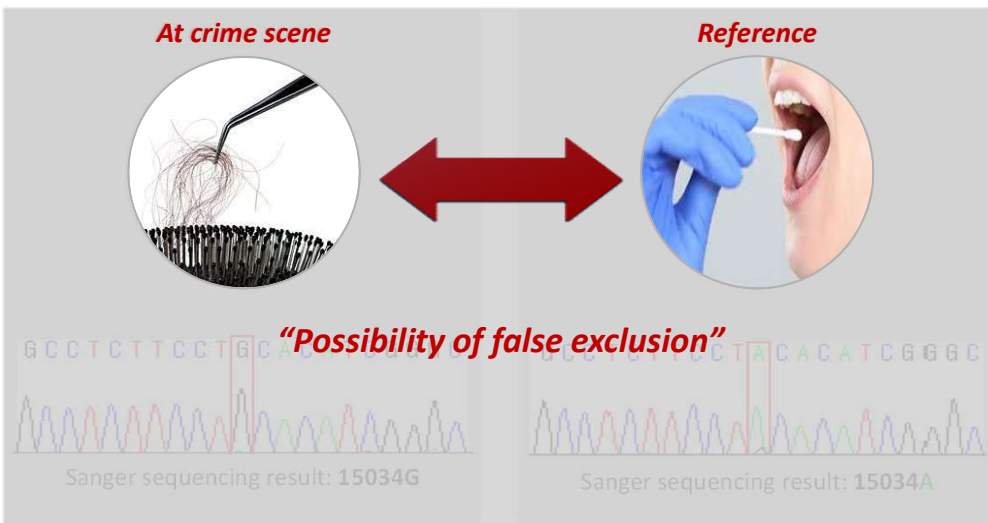
[Sanger sequencing results of *Sample 015*]



4. Discussions

What if..?

[Sanger sequencing results of *Sample 015*]



4. Discussions

What if..?

[Sanger sequencing results of *Sample 015*]

At crime scene



Sanger sequencing result: **15034G**

PHP threshold: approximately ~20%

Reference



Sanger sequencing result: **15034A**

PHP threshold: approximately ~20%

4. Discussions

What if..?

[Sanger sequencing results of *Sample 015*]

At crime scene



Sanger sequencing result: **15034G**



MPS result: **15034G/A_(5.9)**

PHP threshold: 5%

Reference



Sanger sequencing result: **15034A**



MPS result: **15034A/G_(9.9)**

PHP threshold: 5%

5. Conclusions

- We successfully analyzed the whole mtGenome variants of 4 sets of tissue.
- Although **hair shafts** had been stored at room temperature **> 1 year**, their whole mtGenome variants were successfully obtained using the **Precision ID mtDNA Whole Genome Panel**.
- The whole mtGenome variants of **blood and buccal swabs** with 2 **LR-PCR** products were analyzed using the Nextera XT DNA Library Prep Kit.
- In 4 samples, the major nucleotide of PHP was **inverted at one nucleotide** position in the hair shaft and reference samples.
- The MPS data which provide a **high resolution** for PHP, **did not result in exclusion of that both haplotypes were derived from the same donor**.
- **Accumulation of mtGenome PHP data** will facilitate the application of MPS in the comparative analysis of mtGenome variants containing PHP to forensic casework.

5. Conclusions

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ORIGINAL ARTICLE



Comparison of whole mitochondrial genome variants between hair shafts and reference samples using massively parallel sequencing

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