

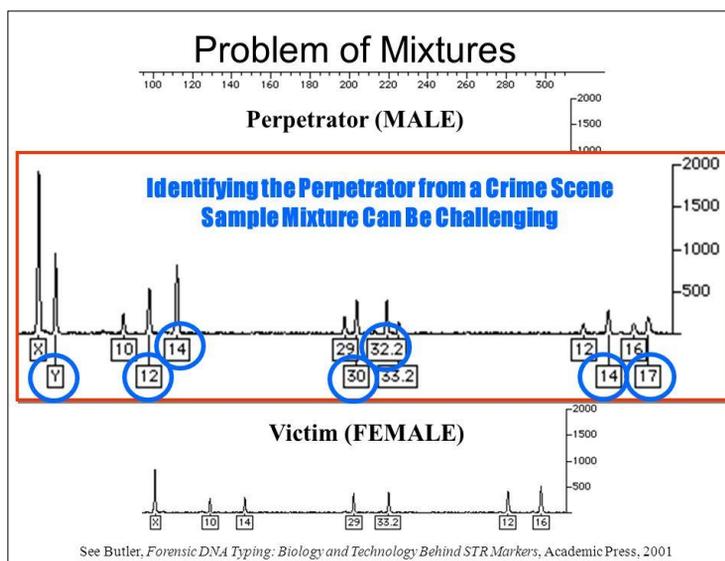


## Microhaplotype

- New marker needed
  - To infer biogeographic ancestry where the STR data is limited
  - To aid in mixture deconvolution and avoid the issue of stutter
- Defined as at least three haplotypes (alleles) composed of multi-SNPs within a region smaller than 200 bp
- NGS enables high-level multiplexing and phase-known haplotyping of microhaplotypes

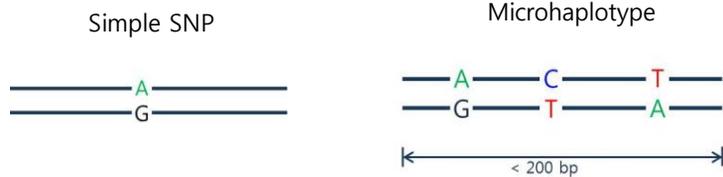
Kidd et al. FSIG, 2014 and 2017

## Background



## Background

### ➤ Simple SNP vs. Microhaplotype



- |                      |                        |
|----------------------|------------------------|
| ▪ Diallelic marker   | ▪ Multi-allelic marker |
| ▪ Low heterozygosity | ▪ High heterozygosity  |
| ▪ ID or ancestry     | ▪ ID and/or ancestry   |

## Aims for this study

- Development of a NGS panel to analyze multiple microhaplotypes
- Reporting statistical value of microhaplotypes in Koreans
- Evaluation of the potential value of microhaplotype in forensics

# Methods

## ➤ Workflow of microhaplotype analysis

1. Target marker selection
2. Primer design to amplify target markers
3. Development of multiplex PCR
4. PCR-based library preparation
5. Library QC & NGS run
6. Data analysis

# Microhaplotype

## ➤ Microhaplotype study

Nearest genes or other symbol	Number of hap alleles		rs-Number dbSNP	Chr	Build 37 nucleotide position	Distance of microhap to base pair
	>10% freq. avg 54 pops	Observed				
CFP104	4	4	rs648344	1	1,741,132	188
USH2A USH2B	4	4	rs660380	1	1,741,139	
			rs6328199	1	21,443,448	18
			rs6604396	1	21,443,445	

This spreadsheet contains all 129 microhaps that have been typed on 55 populations and haplotyped statistically. We have used Informativeness of Rosenberg as a measure of ancestry (L<sub>n</sub>) and Global Average Effective Number of Alleles (A<sub>e</sub>) (InvestGen 2015) as a standardized measure of variation. It is obvious that some microhaplotypes rank very low by both existing statistics. However, we note them in case other populations may change the rankings and/or full sequencing may reveal additional variation making them useful in some situations. Also, some of the microhaplotypes are very close molecularly and have been kept in this listing pending future data than may indicate one of them is significantly better than the other(s). A couple of SNPs occur in overlapping microhaplotypes because we have arbitrarily limited a microhaplotype to an extent of 200 basepairs.

Standardized name using POSIX characters	Global L <sub>n</sub>	Global A <sub>e</sub>	L <sub>n</sub> rank	A <sub>e</sub> rank	chr	p-most chr_position Build GRCh37/hg19	SNPs_Included	KiddGene use_at own_peril	# SNPs
mh13KK-218	0.37857	5.54820	6	1	13	54060827	rs1927847/rs9536429/rs7492234/rs9536430	D13S169	4
mh05KK-170	0.41105	4.89784	4	2	5	2448024	rs74865590/rs438055/rs370672/rs6555108	D5S1970	4
mh21KK-315	0.30144	4.76321	14	3	21	21880086	rs8126597/rs6517970/rs8131148/rs6517971	D21S1263	4
mh21KK-320	0.26139	4.39290	27	4	21	43062859	rs2838081/rs2838082/rs78902658/rs2838083	LINC00111	4
mh11KK-180	0.26272	4.00795	25	5	11	1690791	rs12802112/rs28631755/rs7112918/rs4752777	FAM99A	4
mh13KK-217	0.27577	3.78958	20	6	13	46865930	rs7320507/rs9562648/rs9562649/rs2765614	LRRC63	4
mh01KK-117	0.29949	3.70283	15	7	1	204633340	rs17413714/rs2772234/rs1610401/rs1610400	LRN2	4
mh13KK-223	0.24080	3.66781	34	8	13	110806699	rs1192204/rs1192205/rs3825483/rs3825482	COL4A1	4
mh01KK-205	0.15142	3.65696	85	9	1	18722692	rs11810587/rs1336130/rs1533623/rs1533622	IGSF21	4
mh19KK-299	0.25782	3.65057	28	10	19	22729551	rs4932999/rs4932769/rs2361019/rs280462	LINC01233	4
mh21KK-324	0.14930	3.63751	2	11	21	46714549	rs6518223/rs2838868/rs2729250/rs8133697	LOC642852	4
mh20KK-136	0.21666	3.57664	49	12	2	23809239	rs6714835/rs6756898/rs12617010	COL4A3	3
mh13KK-213	0.22618	3.56103	45	13	13	23765541	rs18181845/rs679482/rs9510616	SCG3	3
mh16KK-255	0.22448	3.45043	46	14	16	81970353	rs18956011/rs3934955/rs3934956/rs4073828	PLCG2	4
mh02KK-134	0.29594	3.35710	17	15	2	161079435	rs93101043/rs3111390/rs72623112	ITGB6	3
mh20KK-307	0.24022	3.28681	35	16	20	16513260	rs6044080/rs17674942/rs6044081/rs16997830	KIF10B	4
mh18KK-293	0.33334	3.24533	8	17	18	76089886	rs621320/rs621340/rs678179/rs621766	D18S1122	4
mh22KK-061	0.20551	3.19248	59	18	22	44763606	rs763040/rs5764924/rs763041	D22S1159	3
mh10KK-169	0.29695	3.18111	16	19	10	14208510	rs10796164/rs10796165/rs17154765/rs10796166	FRMD4A	4
mh09KK-157	0.18911	3.16004	65	20	9	135862479	rs606141/rs8193001/rs56256724/rs633153	GF11B	4
mh03KK-150	0.20618	3.08902	58	21	3	131645972	rs1225051/rs1225050/rs1225049/rs1225048	CPNE4	4
mh11KK-101	0.24528	2.97384	32	22	11	98880163	rs12421109/rs12288401/rs1242081/rs12288402	CNTN5	4

Here we globally

<http://medicine.yale.edu/lab/kidd/publications/publications.aspx>

## Design of multiplex PCR systems

### ◆ Criteria

- Target marker
  - Microhaplotypes suggested by Kidd et al., 2014 and 2015
- Small sized amplicons is adapted as possible
  - while primer is not overlapping with core SNP region
  - finally to be ranged in 90bp ~ 270bp
- Avoid SNP with  $\geq 1\%$  variation reported in primer binding area

### ◆ Resource

- UCSC genome browser (<http://genome.ucsc.edu/>)
- Primer 3 v.0.4.0 (<http://frodo.wi.mit.edu/primer3/>)

## Microhaplotype NGS panel

### ➤ Single tube multiplex PCR system for 51 microhaplotypes

### ➤ Amplicon sizes of 51 microhaplotypes (KplexSeq-MH51)

Group	Marker	Target size (bp)	Group	Marker	Target size (bp)	Group	Marker	Target size (bp)	Group	Marker	Target size (bp)
1	TAS2R1	96	2	CCR2	98	3	TTC12	99	4	DRD3	115
1	USH2A	124	2	PAPD7	130	3	COMT	137	4	ACN9	146
1	COG2	151	2	LINC01233	156	3	GNGT2	157	4	ITGB6	158
1	EDAR	166	2	COL4A3	167	3	CDH4	166	4	D18S1122	169
1	NPEPPS	169	2	SUDS3	180	3	TYRP1	184	4	GFI1B	195
1	D13S169	193	2	D21S1263	198	3	GATA4	200	4	D5S1970	210
1	PLCG2	205	2	D22S1159	216	3	ATXN	220	4	LOC642852	220
1	IGSF21	227	2	KIF16B	221	3	COL4A1	226	4	ADH7	225
1	LRRC	221	2	C14ORF43	230	3	RXRA	233	4	CEP104	232
1	NCAM1	231	2	FAM99A	236	3	SGCG	236	4	FRMD4A	239
1	LINC0111	240	2	OR52S1P	240	3	DLEU2	244	4	LRRN2	246
1	ARHGAP27	249	2	CPNE4	247/252	3	LRRC63	254	4	LYPD6B	250
1	PLIN3	269				3	PAH	270	4	FAT1	268

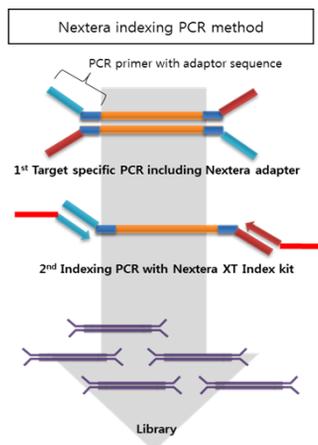
### ➤ DNA samples

- 1ng of 2800M Standard DNA and 122 Korean samples

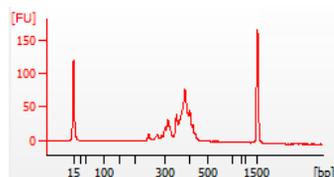
## Methods

### Library preparation

- PCR-based library preparation



- Assess the barcoded library



- Quantify the barcoded library using KAPA library quantification kit

## Two-step PCR for MPS library preparation

- The first PCR using Target specific primers

PCR component	Volume
dH <sub>2</sub> O	2.18
Gold ST*R 10× Buffer	2.0
3× Primer Mix-I & II	6.66
3× Primer Mix-III & IV	6.66
AmpliTaq Gold (5U/uL)	1.5
Template DNA (1ng/uL)	1.0
Total	20.0

- Thermal Cycle

95°C	11 min	} × 26 cycles
94°C	20 sec	
59°C	60 sec	
72°C	45 sec	
72°C	5 min	
4°C	forever	

- The second PCR using Nextera XT Index primers

PCR component	Volume
dH <sub>2</sub> O	12.3
Gold ST*R 10× Buffer	2.0
Index 1 (i5)	2.0
Index 2 (i7)	2.0
AmpliTaq Gold (5U/uL)	0.7
1/10 diluted PCR product	1.0
Total	20.0

- Thermal Cycle

95°C	15 min	} × 15 cycles
94°C	20 sec	
61°C	30 sec	
72°C	45 sec	
72°C	5 min	
4°C	forever	

## Methods

### ➤ Library validation



## Methods

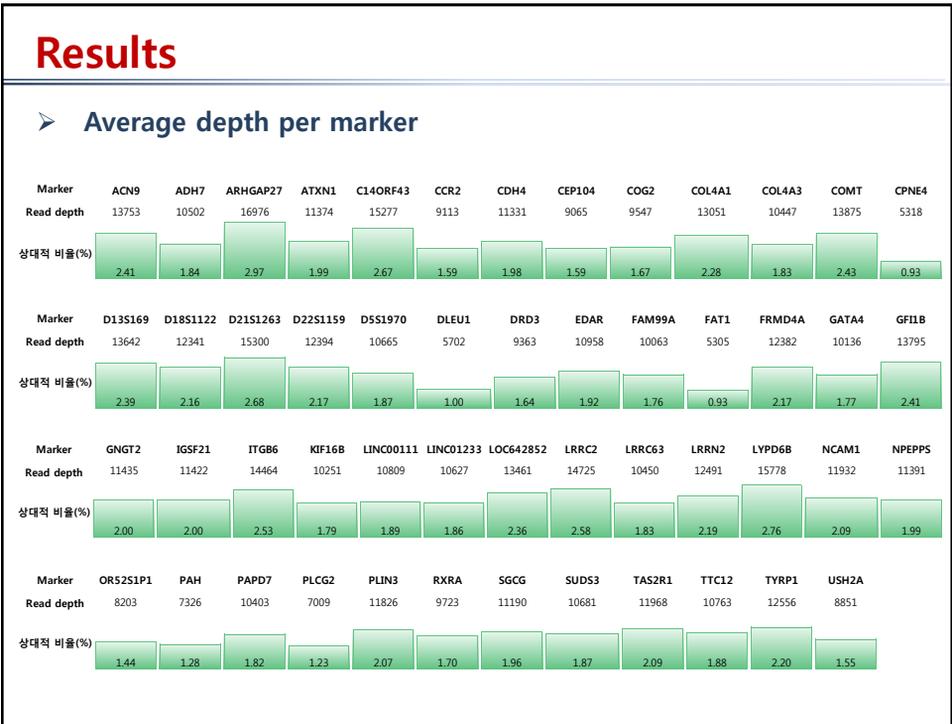
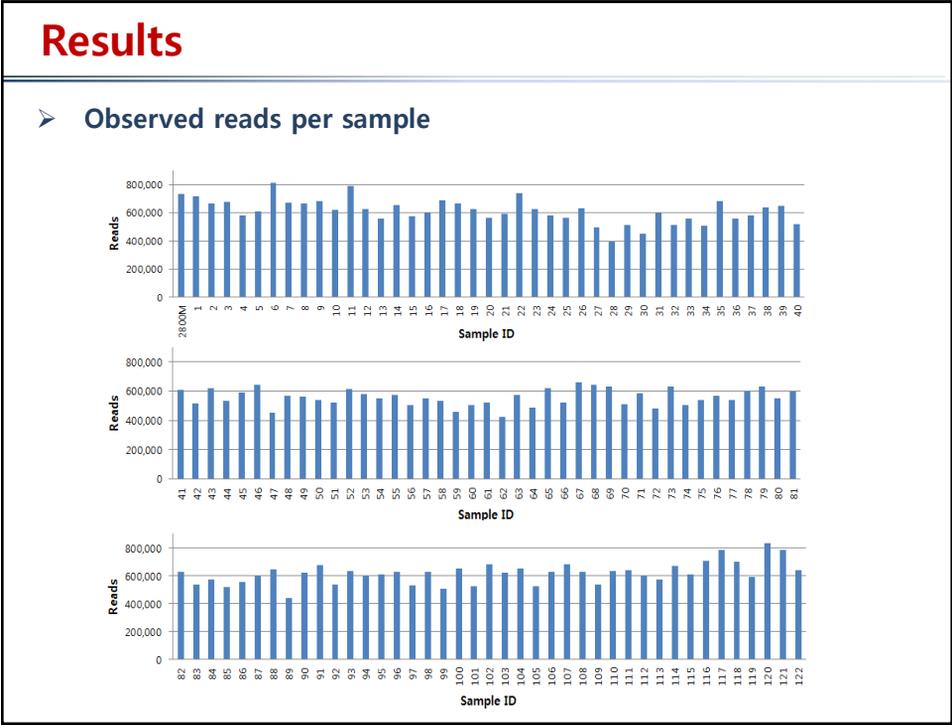
### ➤ NGS run

- Library pooling; final conc. 10 nM
- NGS run on an MiSeq system (Illumina)
- MiSeq Reagent Kit v3, 600 Cycles (2x300 bp)



### ➤ Data analysis

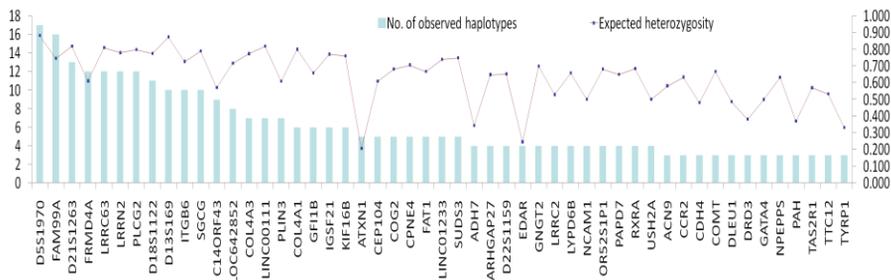
- STRait Razor v2.0 and in-house sequence parsing program
- Data analysis supported by Yonsei Genome Center





## Results

### ➤ Observed haplotypes & frequencies of 51 MHs in Koreans



## Results

Preparing manuscript...

### ➤ Forensic statistical parameters for 51 MHs in Koreans

	H <sub>obs</sub>	H <sub>exp</sub>	Forensic Statistics			Paternity Statistics	
			MP	PD	PIC	PE	TPI
D13S169	0.869	0.875	0.035	0.965	0.857	0.732	3.81
D5S1970	0.885	0.870	0.037	0.963	0.852	0.765	4.36
D21S1263	0.811	0.818	0.060	0.940	0.793	0.621	2.65
LINC00111	0.811	0.819	0.067	0.933	0.790	0.621	2.65
COL4A1	0.779	0.801	0.074	0.926	0.767	0.560	2.26
PLCG2	0.779	0.783	0.080	0.920	0.747	0.560	2.26
LRRN2	0.828	0.780	0.083	0.917	0.748	0.652	2.90
LRRC63	0.787	0.769	0.093	0.907	0.730	0.575	2.35
COL4A3	0.811					0.621	2.65
KF16B	0.697					0.423	1.65
IGSF21	0.664					0.375	1.49
⋮							
CDH4	0.525					0.210	1.05
CEP104	0.549	0.525	0.345	0.655	0.416	0.234	1.11
LRRC2	0.426	0.475	0.370	0.630	0.365	0.131	0.87
PAH	0.402	0.369	0.427	0.573	0.334	0.115	0.84
DRD3	0.385	0.380	0.440	0.560	0.326	0.105	0.81
ADH7	0.344	0.343	0.456	0.544	0.316	0.083	0.76
TYRP1	0.344	0.332	0.492	0.508	0.283	0.083	0.76
EDAR	0.246	0.245	0.592	0.408	0.233	0.044	0.66
A1XN1	0.074	0.103	0.832	0.168	0.100	0.005	0.54

**Match Probability**  
 $= 3.82 \times 10^{37}$

## Summary

- By investigation of 51 microhaplotypes, a total of 196 SNPs and 316 haplotype alleles were observed from 122 Koreans.
- Each microhaplotype consisted of 2 to 7 SNPs, and exhibited 3 to 17 haplotype alleles.
- Overall match probability ( $3.98 \times 10^{39}$ ) of the 51 microhaplotypes was much lower than that of the expanded autosomal STRs.

## Ongoing work

Forensic Science International: Genetics 29 (2017) 29–37

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Contents lists available at ScienceDirect



**Forensic Science International: Genetics**

journal homepage: [www.elsevier.com/locate/fsig](http://www.elsevier.com/locate/fsig)

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Research paper

**Evaluating 130 microhaplotypes across a global set of 83 populations**

Kenneth K. Kidd<sup>a,\*</sup>, William C. Speed<sup>a</sup>, Andrew J. Pakstis<sup>a</sup>, Daniele S. Podini<sup>b</sup>,  
Robert Lagacé<sup>c</sup>, Joseph Chang<sup>c</sup>, Sharon Wootton<sup>c</sup>, Eva Haigh<sup>a</sup>, Usha Soundararajan<sup>a</sup>

<sup>a</sup> Department of Genetics, Yale University School of Medicine, 333 Cedar Street, New Haven, CT, 06520-8005, USA  
<sup>b</sup> Department of Forensic Sciences, The George Washington University, 2100 Foxhall Road, NW, Washington, D.C., 20007, USA  
<sup>c</sup> Human Identification Group, ThermoFisher Scientific, 180 Oyster Point Blvd, South San Francisco, CA, 94080, USA

- **Developing modified NGS panel to analyze microhaplotypes ...**

## Future work

- Resolution in mixture deconvolution
  - Mixed DNA with ratio of 1:1, 1:3, 1:6, 1:9, 1:19, 1:29, 1:50 and 1:100
- Evaluation of biogeographic inference
  - DNA from 4 population including Koreans
  - Adapted new informative microhaplotypes
- Application to complex kinship case

## Acknowledgements

### ❖ Yonsei DNA profiling group

Eun Young Lee, MS  
So Yeun Kwon, MS  
Sang Eun Jung, MS  
Hwan Young Lee, Ph.D.



### ❖ Yonsei Genome Center

Sora Kim, Ph.D. candidate  
Sang Woo Kim, Ph.D.

➤ This work was supported by a fund (No. NFS2016DNA02) from the Forensic Research Program of the National Forensic Service, Korea.